Matthew W. Fields

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Education and Training

Postdoctoral Research Associate, Oak Ridge National Laboratory	<u>2001</u>
Ph.D. Microbiology, Minor: Biochemistry/Biological Engineering, Cornell University	<u>2001</u>
M.S. Biological Sciences, Mississippi State University	<u> 1995</u>
B.S. Biology/Chemistry, Western Kentucky University	<u>1993</u>

Research Experience

Assistant Professor, Department of Microbiology, Miami University

2003-

Develop and direct research program in bacterial physiology, biochemistry, and ecology dealing with model organisms (*Shewanella* spp., *Desulfovibrio*, *Alkaliphilus*, *Anaeromyxobacter*, field isolates) and *in situ* communities associated with bioremediation (groundwater, sediments, reactors)

Current Laboratory 1 Postdoctoral Assoc.; 3 Ph.D. students; 4 undergraduates

Presentations (oral/poster at national conferences)

17 in 2005; 13 in 2004; 10 in 2003

Research Staff Scientist, Oak Ridge National Laboratory

2001-2003

Assess microbial ecology of contaminated sites using molecular- and culture-based techniques; isolation and physiological characterization of novel and unique microbes; development of functional, community, and metagenomic DNA microarrays; coordination of global transcriptomic studies with cellular physiology

Graduate Research Assistant, Department of Microbiology, Cornell University

Conducted and initiated independent research combining microbial physiology and molecular biology dealing with the role and regulation of polysaccharide hydrolases from rumen bacteria; advised and coordinated undergraduate research

Teaching Experience

Microbial Ecology (475/575) lecture

2005-

The course covered the basic concepts of microbial ecology, examined biotic and abiotic interactions within and between populations, and discussed the role of bacteria in maintaining steady-state conditions in natural ecosystems. Lecture material was supplemented with scientific literature for discussion-based meetings.

Biological Concepts (116) lecture (co-instructor)

2004-

Covered basic prokaryotic and eukaryotic taxonomy, DNA replication, transcription, translation, and biotechnology.

Summer Workshop for Secondary Education Teachers (co-instructor

2004

Lectures covered basic microbiological concepts, practices, and experiments.

Microbial Physiology (425/525) lecture and laboratory

2003-

The course covered principle concepts of microbial physiology including: cell composition, cell growth, electron transport, chemiosmosis, photosynthesis, central metabolism, lipids, nucleic acids, transport, gene expression, nutrient transport, and adaptive response.

Graduate Journal Club (750) course

2003-

Consisted of weekly meetings with graduate students to discuss primary literature on stress response studies related to microbial physiology. Scientific papers were discussed in detail in an informal setting to stimulate open discussion and questions in a round-table format.

University Service

Editorial Board for Applied and Environmental Microbiology	2006-2009
Invited speaker at MI-ASM branch meeting	2005
Invited by DOD to attend MBT workshop and draft strategic plan	2005
Faculty Research Committee (advisory committee to University Senate)	2005-
Invited by USDA to help develop road-map for a joint NSF-USDA proposal call	2004
Ad Hoc reviewer for Applied and Environmental Microbiology	2004-
Serve on the Ecology program committee; Instrumentation Laboratory Advisory Committee	2004-
Serve as MBI advisor for Environmental Science co-major	2004-
Faculty Fellow for the Sigma Phi Epsilon fraternity	2004-
Co-PI for funded NSF equipment grant	2004-

Publications

- 27. Hwang, C., W.-M.Wu, T.J. Gentry, J. Carley, S.L. Carroll, C. Schadt, D. Watson, P.M. Jardine, J. Zhou, R.F. Hickey, C.S. Criddle, and M.W. Fields. 2005. Changes in bacterial community structure correlate with initial operating conditions of a field-scale denitrifying fluidized bed reactor. Appl. Microbiol. Biotech. (accepted)
- **26.** Fields, M.W., J. Schryver, C.C. Brandt, T. Yan, J. Zhou, and A.V. Palumbo. 2005. Confidence intervals for similarity values achieved from direct sequence determination of cloned SSU rRNA genes from environmental samples. J. Microbiol. Methods (in press)
- 25. M. Gentile, T. Yan, S.M. Tiquia, M.W. Fields, J. Nyman, J. Zhou, and C. S. Criddle. 2005. Stability in a denitrifying fluidized bed reactor. Microbial Ecology (in press)
- 24. B. Gu, W.-M. Wu, M.A. Ginder-Vogel, H. Yan, M.W. Fields, S. Fendorf, C.S. Criddle, and P.M. Jardine. 2005. Bioreduction of uranium in a contaminated soil column. Environ. Science. Tech. 39:4841-4847
- 23. Dong, H., H. Jiang, G. Zhang, B. Yu, L.R. Chapman, C.R. Lucas, and M.W. Fields. 2005. Microbial diversity in water and sediment of Lake Qinghai: the largest inland saline lake in China. Microbial Ecol. (in press)
- 22. He, Z., L. Wu, M.W. Fields and J. Zhou. 2005. Comparison of microarrays with different probe sizes for monitoring gene expression. Appl. Environ. Microbiol. 71:5154-5162
- **21.** Fields, M.W., T. Yan, S.-K. Rhee, S.L. Carroll, P.M. Jardine, D.B. Watson, C.S. Criddle and J. Zhou. 2005. Impacts on microbial communities and cultivable isolates from groundwater contaminated with high levels of nitric acid-uranium waste. FEMS Microboil. Ecol. 53:417-428
- 20. Wu, W., B. Gu, M.W. Fields, M. Gentile, S.M. Tiquia, J. Nyman, J. Zhou, C.S. Criddle. 2005. Characterization of uranium (VI) reduction by microbial biomass from a denitrifying fluidized bed reactor. Bioremediation J. 9.1-13
- 19. Fields, M.W. and J.B. Russell. 2005. Transcriptional regulation of β -glucanase activity in the ruminal bacterium, *Prevotella bryantii* B₁4. Curr. Microbiol. 50:155-159
- 18. He, Z., L. Wu, X. Li, M.W. Fields and J. Zhou. 2005. Empirical establishment of oligonucleotide probe design criteria. Appl. Environ. Microbiol. 71:3753-3760
- 17. Liu, Y., W. Gao, L. Wu, X. Liu, T. Yan, M.W. Fields, D.K. Thompson, and J. Zhou. 2005.

- Genomic expression profiling of *Shewanella oneidensis* MR-1 response to sodium salt stress. J. Bacteriol. 187:2501-2507
- **16.** Palumbo, A.V., J.C. Schryver, M.W. Fields, C. E Bagwell, J. Zhou, T. Yan, X. Liu and C.C. Brandt. 2004. Coupling functional gene diversity and geochemical data from environmental samples. Appl. Environ. Microbiol. 70:6525-6534
- 15. Tiquia, S.M., S.C. Chong, M.W. Fields, and J. Zhou. 2004. Oligonucleotide-based functional gene arrays for analysis of microbial communities in the environment, *In* Molecular Microbial Ecology Manual, Kluwer Academic Press
- 14. Wu, L., D.K. Thompson, X. Liu, M.W. Fields, C.E. Bagwell, J.M. Tiedje, and J. Zhou. 2004. Development and evaluation of microarray-based whole-genome hybridization for detection of microorganisms within the context of environmental applications. Environ. Sci. Technol. 38:6775-6782
- 13. Ye, Q., Y.Roh, B.B. Blair, C. Zhang, J. Zhou and M.W. Fields. 2004. Isolation and characterization of a novel, alkaliphilic, metal-reducing bacterium, and possible implications for alkaline chemotrophy. Appl. Environ. Microbiol. 70:5595-5602
- 12. Yan, T.^, M.W. Fields^, L. Wu, Y. Zu, J.M. Tiedje, and J. Zhou. 2003. Molecular diversity and characterization of nitrite reductase gene fragments (*nir*S and *nir*K) from nitrate- and uranium-contaminated groundwater. Environ. Microbiol. 5(1):13-24
- 11. Fields, M.W. and J.B. Russell. 2002. The glucomannokinase of *Prevotella bryantii* B₁4: sequence homology with the regulatory glucokinases of gram-positive microorganisms. Anaerobe 8:69-74
- 10. A.S. Beliaev, D.K. Thompson, M.W. Fields, L.Wu, D.P. Lies, K.H. Nealson, and J. Zhou. 2002. Microarray transcription profiling of a *Shewanella oneidensis etr* A mutant. J. Bacteriol. 184:4612-16
- 9. Roh, Y., R. Stapleton, J. Zhou, C. Zhang, A.V. Palumbo, T.J. Phelps, and M.W. Fields. 2002. Iron reduction by extremophiles: Geochemical and biotechnological implications, *In* The Biogeochemistry of Iron Cycling in the Environment, C. Zhang and J. D. Coates (eds.). Kluwer Academic Press
- 8. Fields, M.W. and J.B. Russell. 2001. Purification and characterization of a novel glucomannokinase of *Prevotella bryantii* B_14 , and its possible role in β -glucanase expression. Microbiol. 147:1035-1043
- 7. Jarvis, G.N., M.W. Fields, D.A. Adamovich, C.E. Arthurs, and J.B. Russell. 2001. The mechanism of carbonate killing of *Escherichia coli*. Letts. Appl. Microbiol. 33:196-200
- **6.** Fields, M.W., S. Mallik, and J.B. Russell. 2000. Fibrobacter succinogenes S85 ferments cellulose just as fast as cellobiose until limited by cellulose surface area. Appl. Microbiol. Biotech. 54:570-574
- **5.** Fields, M.W. and J.B. Russell. 2000. Alternative pathways of glucose transport in *Prevotella bryantii* B₁4. FEMS Microbiol. Letts. 183:137-142
- 4. Fields, M.W. and J.B. Russell. 1998. The role of ruminal carboxymethylcellulases in the degradation of β -glucans from cereal grains. FEMS Microbiol. Ecol. 27:261-268

- 3. Fields, M.W., D.B. Wilson, and J.B. Russell. 1997. A mutant of *Prevotella ruminicola* B₁4 deficient in β-1,4-endoglucanase and mannanase activities. FEMS Microbiol. Letts. 154:9-15
- **2.** Gardner, R.G., J.E. Wells, **M.W. Fields**, D.B. Wilson, and J.B. Russell. 1997. A *Prevotella ruminicola* B₁4 operon encoding extracellular polysaccharide hydrolases. Curr. Microbiol. 35:274-277
- 1. Fields, M.W., P.E. Ryals, and K.L. Anderson. 1997. Polysaccharide-inducible outer membrane proteins of *Bacteroides xylanolyticus* X5-1. Anaerobe 3:43-48

Manuscripts

- H. Jiang, G. Zhang, L. Chapman, B. Yu, M.W. Fields, and H. Dong. Microbial diversity in water and sediment of lake chaka: an inland hypersaline lake in northwestern china. Appl. Environ. Microbiol. (in revision)
- Nowlin, W.H., M.J. González, M.H.H. Stevens, M.J. Vanni, M.W. Fields, J.J. Valente. Periodical cicadas affect dynamics, productivity and stability of woodland pond ecosystems. Ecol. Letts (submitted)
- Kurowski, J., A. Sundararaja, A. Klonowska, T. Yan D.M. Klingman, M. Duley, J. Zhou, and M. W. Fields. Deletion of a multi-domain PAS protein causes pleiotropic effects in *Shewanella oneidensis* MR-1. J. Bacteriol. (in review)
- **Fields, M.W.**, C.E. Bagwell, S.L. Carroll, T. Yan, X. Liu, D.B. Watson, P.M. Jaridne, C.S.Criddle, T.C. Hazen, and J. Zhou. Gene sequences as indicators of bacterial community responses to mixed-waste contamination. Environ. Sci. Technol. (in review)
- He, Q. et al. Energetic consequences of nitrite stress in *Desulfovibrio vulgaris* Hildenborough inferred from global transcriptional analysis. Mol. Microbiol. (submitted)
- Wu, W. et al. Field-scale bioremedation of uranium in a highly contaminated aquifer II: reduction of U(VI) and geochemical control of U(VI) bioavailability. Environ. Sci. Technol. (submitted)
- Clark, M.E., Q. He, Z. He, E. Alm, K. Huang, T.C. Hazen, A. Arkin, J.D. Wall, J. Zhou, and M.W. Fields. Temporal transcriptomic analyses of *Desulfovibrio vulgaris* Hildenborough grown under electron acceptor excess conditions. J. Bacteriol. (in preparation)

Current Grant Support

- PI Construction of whole genome microarrays, and expression analysis of *Desulfovibrio vulgaris* cells in metal-reducing conditions (uranium and chromium), DOE-ER63765
- Co-PI Integrated analysis of protein complexes and regulatory networks involved in anaerobic energy metabolism of *Shewanella oneidensis* MR-1, LAB 01-20, DOE Microbial Cell Project
- Co-PI Rapid deduction of stress response pathways in metal/radionuclide reducing bacteria, LAB 02-13, DOE Genomes to Life program
- Co-PI Effects of 17 year cicada emergence on aquatic food webs, NSF 04-2, DEB-Ecology
- PI Whole-genome sequence determination of novel, extremophilic, metal-reducing bacteria, DOE Community Sequencing Program
- Co-PI Genome-level understanding of the diversity and structure of a groundwater microbial community in the NABIR Research Field Research Center, DOE Community Sequencing Program
- PI Identification of Molecular and Cellular Responses of *Desulfovibrio vulgaris* Biofilms under Culture Conditions Relevant to Field Conditions for Bioreduction of Heavy Metals, DOE-DE-FG01-05 ER05-05